EAGLE Environmental, Inc.

- Industrial Hygiene / IAQ
- Hazardous Building Materials
- Environmental Assessments
- Laboratory Services & Training

March 26, 2025

Mr. Joseph Tommaselli
Buildings and Grounds Supervisor
Area Cooperative Educational Services (ACES)
350 State Street
North Haven, Connecticut 06473

RE: Targeted Post Water Intrusion Mold & Moisture Assessment
Wintergreen Interdistrict Magnet School – First Floor Corridor
670 Wintergreen Avenue
Hamden, Connecticut
Eagle Project No. 25-064.11T1

Dear Mr. Tommaselli:

Eagle Environmental, Inc (Eagle) conducted a Targeted Post Water Intrusion Mold and Moisture Assessment (Assessment) at the first-floor corridor of the Wintergreen Interdistrict Magnet School located at 670 Wintergreen Avenue in Hamden, Connecticut (Site). The Assessment was conducted to verify the efficacy of post water intrusion cleaning and drying activities within the affected areas of the first-floor corridor at the Site building.

Please do not hesitate to contact us if you have any questions regarding the contents of this report.

Sincerely,
Eagle Environmental, Inc.

Report Prepared By:
Jasone Eberhard
Senior Project Manager

Z:\2025 Files\2025 Reports\ACES\WIMS\Corridor Water Intrusions Targeted Mold & Moisture Assmt & Airborne Particulate Sampling.Docx

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1.0 INTRODUCTION

1.1 Background

Eagle Environmental, Inc (Eagle) conducted a Targeted Post Water Intrusion Mold and Moisture Assessment (Assessment) at the first-floor corridor of the Wintergreen Interdistrict Magnet School located at 670 Wintergreen Avenue in Hamden, Connecticut (Site). The Assessment was conducted to verify the efficacy of post water intrusion cleaning and drying activities within the affected areas of the first-floor corridor at the Site building.

It was reported that the Site had previously experienced a water loss within the first-floor corridor. The water intrusion was reported to have been corrected and carpeted floor surfaces were extracted and disinfected prior to Eagle' Assessment. The scope of Eagle's services included a physical inspection for mold and moisture, moisture mapping and non-viable spore trap (air) sampling.

The Assessment was conducted on March 12, 2024 by Jonathan Vargas, Environmental Consultant II for Eagle.

1.2 Objectives and Limitations

The Assessment included the following tasks:

1. A physical inspection to evaluate the targeted area of the Site with regard to mold and moisture;
2. Moisture mapping;
3. Non-viable spore trap (air) sampling to characterize the targeted area of the Site with regard to fungal ecology; and,

Observations and data obtained during the Assessment represent conditions during that time span only. Fungal ecology and psychrometric variables are intrinsically dynamic and can vary substantially.

2.0 ASSESSMENT AND SAMPLING METHODOLOGY

2.1 Physical Inspection

Eagle performed a physical inspection of the affected areas of the first-floor corridor as indicated by school staff. Visible accessible surfaces of interior building finishes, furnishings, and stored contents were examined for indications of suspect visible mold growth and/or moisture impact. Musty, moldy, or other malodors were recorded if they were encountered.

2.2 Moisture Mapping

Eagle used a Fluke Ti360+ Thermal Imager to scan building finishes and furnishings for indications of elevated moisture. The thermal imager uses infra-red technology to display locations of surface temperature differential which could potentially indicate elevated moisture levels in building materials.

Eagle used a Protimeter Survey Master Moisture Measurement System moisture meter fitted with an appropriate electrode to measure the moisture content in potentially wet locations.

2.3 Spore Trap (Air) Sampling

Presently there are no established regulatory standards for airborne mold spore concentrations.

Eagle conducted non-viable fungal particulate air sampling in the targeted area of the Site to determine airborne levels of non-viable fungal particulates. Additionally, a sample was collected from an outdoor location for comparison purposes.

The characterization of the indoor air with regard to fungal spore concentrations was intended to screen the Site for excessive fungal spore concentrations. The spore concentrations reported are analyzed as numerical indicators and do not represent statistically valid hard numbers.

Airborne spore concentrations that are significantly elevated above background concentrations suggest the presence of active or sporulating mold colonies. Generally, and with consideration to site-specific conditions, Eagle considers mold spore concentrations in excess of one thousand (1000) Counts/m^3 per genera indicative of a potential IICRC Condition 2 (settled spores) or Condition 3 (fungal growth) when significantly (generally an order of magnitude) in excess of the corresponding background genera.

Non-viable fungal air sampling was performed using Air-O-Cell spore trap cassettes for total spore count analysis. The spore-trap cassettes were attached to a Buck BioAir Pump to capture airborne mold spores. The sampling period for each air sample was ten (10) minutes at a flow rate of fifteen (15) liters per minute for a total volume of one-hundred and fifty (150) liters. The pump was calibrated with a calibration standard at the beginning and end of each sampling period.

The samples were labeled and placed into a sealed plastic bag for transport to EMLab P&K in Marlton, New Jersey. EMLab P&K is accredited by the American Industrial Hygiene Association (AIHA) for mold analysis and participates in the Environmental Microbiology Laboratory Accreditation Program (EMLAP).

3.0 ASSESSMENT AND SAMPLING FINDINGS

3.1 Physical Inspection Findings

The affected areas of the corridor, as indicated by school staff, were observed to be clean. No malodors were encountered and no visible suspect mold was observed on accessible surfaces.

3.2 Moisture Mapping Findings

IR camera scanning and follow up moisture meter testing identified one area of carpeting within the corridor at Room 219 as being in an "at risk" condition at the time of the Assessment. Eagle was unable to verify if the area of "at risk" condition was directly attributable to the water intrusion event.

3.3 Spore Trap (Air) Sampling Findings

Eagle collected three (3) air samples from the targeted area at the Site. One (1) air sample was collected outside of the building for use as a baseline comparison sample. Four (4) samples were submitted to the laboratory for analyses.

Summary of Analytical Results Table:

Sample Number	Location	Fungal Genera	Spore Concentration/m^3	Total Spore Concentration
A1	North Side of Affected Corridor Area	*Basidiospores*	160	220
		Smuts, Periconia, Myxomycetes	60	
A2	Center of Affected Corridor Area	*Basidiospores*	110	180
		Cladosporium	53	
		Smuts, Periconia, Myxomycetes	20	
A3	South Side of Affected Corridor Area	*Basidiospores*	210	270
		Smuts, Periconia, Myxomycetes	60	
A4	Exterior	*Cladosporium*	80	80

Spore concentrations are reported in spores/cubic meter of air. Total spores/cubic meter are rounded to two significant figures to reflect analytical precision.

Overall spore concentrations on the interior air samples were above the exterior baseline air sample at the time of the sampling. *Basidiospores* and *Smuts, Periconia, Myxomycetes* type spores were detected on the interior air samples. These spore types were not present on the exterior baseline sample.

Due to the presence of *Basidiospores* and *Smuts, Periconia, Myxomycetes* species type spores on the interior air samples, Eagle requested the laboratory perform a MoldSCORE™ analysis of the air samples. The MoldSCORE is a statistical calculation of the likelihood of the spores present on an indoor sample originating from the interior of the building due to active mold growth. MoldSCOREs are rated on a scale from 100 to 300. MoldSCOREs below 150 represent a low probability that the mold spores originated from the interior of the building. MoldSCOREs above 250 represent a high likelihood that the spores originated from inside the building, presumably from indoor mold growth. Mold scores between 150 and 250 indicate a moderate likelihood of indoor mold growth. The representative MoldSCORE for each of the tested locations is given below:

Location	MoldSCORE
North Side of Affected Corridor Area	117
Center of Affected Corridor Area	112
South Side of Affected Corridor Area	122

The MoldSCOREs for the three (3) air samples collected in the first-floor corridor were below 150, indicating a low probability that these spores originated from the interior of the building. Based upon the air sample results, the targeted area displayed an IICRC Condition 1 – "Normal Fungal Ecology" at the time of the Assessment.

The Mold Laboratory Report and Chain of Custody are included as Appendix A. The Laboratory Certificate of Accreditation is included as Appendix B.

4.0 CONCLUSIONS AND RECOMMENDATIONS

4.1 Conclusions

Based upon the physical inspection, moisture mapping, and sampling data, it is concluded that a normal fungal ecology was displayed at the targeted areas at the time of the post water intrusion Assessment.

Observations and data obtained during the Assessment represent conditions during that time span only. Fungal ecology and psychrometric variables are intrinsically dynamic and can vary substantially.

4.2 Recommendations

Eagle recommends that additional carpet drying be conducted in the corridor at Room 219 based upon the indications of an "at risk" area of carpeting at this location.

4.3 General Recommendations for Mold and Moisture Remediation

These recommendations are time sensitive and subject to change due to the variability of fungal ecologies, atmospheric conditions and building dynamics.

All mold and water remediation work should be performed in accordance with the recommendations of the Institute of Inspection, Cleaning and Restoration Certification (IICRC) S-500, Standard and Reference Guide for Professional Water Damage Restoration – Third (or most recent) Edition, the IICRC S-500, Standard and Reference Guide for Professional Mold Remediation – Second (or most recent) Edition, the New York City Department of Health and Mental Hygiene Guidelines on Assessment and Remediation on Fungi in Indoor Environments (issued November, 2008) and the Connecticut Department of Public Health, Guidance for Mold Abatement Contractors.

Disinfectants used in cleaning should be compatible with the building and building occupants and should be listed in the Environmental Protection Agency (EPA) Federal Insecticide, Fungicide and Rodenticide FIFRA database and used in strict accordance with the Manufacturer's recommendations.

APPENDIX A

MOLD LABORATORY REPORTS

eurofins | Built Environment Testing

Report for:

8 S Main Street
Terryville, CT 06786

Regarding: Eurofins Built Environment Testing East, LLC
Project: 25 - oh4.1171 ALES; 670 Wintergreen Ave
EML ID: 3987863

Approved by:

Technical Manager
Ariunaa Jalsrai

Dates of Analysis:
Spore trap analysis: 03-17-2025

Service SOPs: Spore trap analysis (EB-MY-S-1038)
AIHA LAP, LLC accredited service, Lab ID #103005

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received and tested. Information supplied by the client which can affect the validity of results: sample air volume.

Eurofins Built Environment Testing East, LLC ("the Company"), a member of the Eurofins Built Environment Testing group of companies, shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.
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Eurofins Built Environment Testing East, LLC
3000 Lincoln Drive East, Suite A, Marlton, NJ 08053
(866) 871-1984 www.eurofinsus.com/Built

Client: Eagle Environmental, Inc.
C/O: Mr. Jason Eberhard
Re: 25 - oh4.1171 ALES; 670 Wintergreen Ave

Date of Sampling: 03-12-2025
Date of Receipt: 03-17-2025
Date of Report: 03-17-2025

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	A1: Hallway			A2: Hallway		
Comments (see below)	None			None		
Lab ID-Version‡:	19831675-1			19831676-1		
Analysis Date:	03/17/2025			03/17/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Ascospores						
Basidiospores	6	25	160	4	25	110
Bipolaris/Drechslera group						
Botrytis						
Chaetomium						
Cladosporium				2	25	53
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes	9	100	60	3	100	20
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 7			< 7		
Pollen/m3	< 7			< 7		
Skin cells (1-4+)	2+			2+		
Sample volume (liters)	150			150		
§ TOTAL SPORES/m3			220			180

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium, Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.
‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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Client: Eagle Environmental, Inc.
C/O: Mr. Jason Eberhard
Re: 25 - oh4.1171 ALES; 670 Wintergreen Ave

Date of Sampling: 03-12-2025
Date of Receipt: 03-17-2025
Date of Report: 03-17-2025

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	A3: Hallway			A4: Exterior		
Comments (see below)	None			None		
Lab ID-Version‡:	19831677-1			19831678-1		
Analysis Date:	03/17/2025			03/17/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Ascospores						
Basidiospores	8	25	210			
Bipolaris/Drechslera group						
Botrytis						
Chaetomium						
Cladosporium				3	25	80
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes	9	100	60			
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 7			< 7		
Pollen/m3	< 7			47		
Skin cells (1-4+)	2+			< 1+		
Sample volume (liters)	150			150		
§ TOTAL SPORES/m3			270			80

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium, Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3, per spore and per sample.

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§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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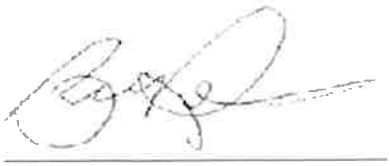
Client: Eagle Environmental, Inc.
C/O: Mr. Jason Eberhard
Re: 25 - oh4.1171 ALES; 670 Wintergreen Ave

Date of Sampling: 03-12-2025
Date of Receipt: 03-17-2025
Date of Report: 03-17-2025

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

PROJECT ANALYST AND SIGNATORY REPORT

Project Analyst



Analyst: Benjamin Reich

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

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Client: Eagle Environmental, Inc.
C/O: Mr. Jason Eberhard
Re: 25 - oh4.1171 ALES; 670 Wintergreen Ave

Date of Sampling: 03-12-2025
Date of Receipt: 03-17-2025
Date of Report: 03-18-2025

MoldSCORE™: Spore Trap Report

Outdoor Sample: A4 Exterior

Fungi Identified	Outdoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3
Generally able to grow indoors*			
Alternaria		ND	< 7
Bipolaris/Drechslera group		ND	< 7
Chaetomium		ND	< 7
Cladosporium		3	80
Curvularia		ND	< 7
Nigrospora		ND	< 7
Penicillium/Aspergillus types†		ND	< 7
Stachybotrys		ND	< 7
Torula		ND	< 7
Seldom found growing indoors**			
Ascospores		ND	< 7
Basidiospores		ND	< 7
Rusts		ND	< 7
Smuts, Periconia, Myxomycetes		ND	< 7
Total			**80**

Location: A1 Hallway

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 7		100
Bipolaris/Drechslera group		ND	< 7		100
Chaetomium		ND	< 7		100
Cladosporium		ND	< 7		100
Curvularia		ND	< 7		100
Nigrospora		ND	< 7		100
Penicillium/Aspergillus types†		ND	< 7		100
Stachybotrys		ND	< 7		100
Torula		ND	< 7		100
Seldom found growing indoors**					
Ascospores		ND	< 7		100
Basidiospores		6	160		117
Rusts		ND	< 7		100
Smuts, Periconia, Myxomycetes		9	60		112
Total			**220**	**Final MoldSCORE**	**117**

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Client: Eagle Environmental, Inc.
C/O: Mr. Jason Eberhard
Re: 25 - oh4.1171 ALES; 670 Wintergreen Ave

Date of Sampling: 03-12-2025
Date of Receipt: 03-17-2025
Date of Report: 03-18-2025

MoldSCORE™: Spore Trap Report

Location: A2 Hallway

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 7		100
Bipolaris/Drechslera group		ND	< 7		100
Chaetomium		ND	< 7		100
Cladosporium		2	53		103
Curvularia		ND	< 7		100
Nigrospora		ND	< 7		100
Penicillium/Aspergillus types†		ND	< 7		100
Stachybotrys		ND	< 7		100
Torula		ND	< 7		100
Seldom found growing indoors**					
Ascospores		ND	< 7		100
Basidiospores		4	110		112
Rusts		ND	< 7		100
Smuts, Periconia, Myxomycetes		3	20		104
Total			**180**	**Final MoldSCORE**	**112**

Location: A3 Hallway

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 7		100
Bipolaris/Drechslera group		ND	< 7		100
Chaetomium		ND	< 7		100
Cladosporium		ND	< 7		100
Curvularia		ND	< 7		100
Nigrospora		ND	< 7		100
Penicillium/Aspergillus types†		ND	< 7		100
Stachybotrys		ND	< 7		100
Torula		ND	< 7		100
Seldom found growing indoors**					
Ascospores		ND	< 7		100
Basidiospores		8	210		122
Rusts		ND	< 7		100
Smuts, Periconia, Myxomycetes		9	60		112
Total			**273**	**Final MoldSCORE**	**122**

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C/O: Mr. Jason Eberhard
Re: 25 - oh4.1171 ALES; 670 Wintergreen Ave

Date of Sampling: 03-12-2025
Date of Receipt: 03-17-2025
Date of Report: 03-18-2025

MoldSCORE™: Spore Trap Report

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

†The spores of Aspergillus and Penicillium (and others such as Acremonium, Paecilomyces) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods.

‡Rated on a scale from 100 to 300. A rating less than 150 is low and indicates a low probability of spores originating inside. A rating greater than 250 is high and indicates a high probability that the spores originated from inside, presumably from indoor mold growth. A rating between 150 and 250 indicates a moderate likelihood of indoor fungal growth. MoldSCORE is NOT intended for wall cavity samples. It is intended for ambient air samples in residences. Using the analysis on other samples (like wall cavity samples) will lead to misleading results.

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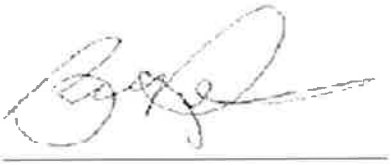
MoldSCORE™: Spore Trap Report

PROJECT ANALYST AND SIGNATORY REPORT

Project Analyst



Analyst: Benjamin Reich

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

APPENDIX B

LABORATORY CERTIFICATES OF ACCREDITATION

AIHA Laboratory Accreditation Programs, LLC

acknowledges that

Eurofins EPK Built Environment Testing - Marlton, NJ

3000 Lincoln Dr E Ste A Marlton, NJ 08053-1500

Laboratory ID: LAP-103005

along with all premises from which key activities are performed, as listed above, has fulfilled the requirements of the AIHA Laboratory Accreditation Programs (AIHA LAP), LLC accreditation to the ISO/IEC 17025:2017 international standard, General Requirements for the Competence of Testing and Calibration Laboratories in the following:

LABORATORY ACCREDITATION PROGRAMS

☐	**INDUSTRIAL HYGIENE**	Accreditation Expires:
☐	**ENVIRONMENTAL LEAD**	Accreditation Expires:
☑	**ENVIRONMENTAL MICROBIOLOGY**	Accreditation Expires: June 01, 2025
☐	**FOOD**	Accreditation Expires:
☐	**UNIQUE SCOPES**	Accreditation Expires:

Specific Field(s) of Testing (FoT)/Method(s) within each Accreditation Program for which the above named laboratory maintains accreditation is outlined on the attached Scope of Accreditation. Continued accreditation is contingent upon successful on-going compliance with ISO/IEC 17025:2017 and AIHA LAP, LLC requirements. This certificate is not valid without the attached Scope of Accreditation. Please review the AIHA LAP, LLC website (www.aihaaccreditedlabs.org) for the most current Scope.

Cheryl O. Morton

Cheryl O Morton
Managing Director, AIHA Laboratory Accreditation Programs, LLC

Revision21: 05/15/2023

Date Issued: 06/01/2023

AIHA Laboratory Accreditation Programs, LLC

SCOPE OF ACCREDITATION

Eurofins EPK Built Environment Testing - Marlton, NJ

3000 Lincoln Dr E Ste A Marlton, NJ 08053-1500

Laboratory ID: LAP-103005

Issue Date: 06/01/2023

The laboratory is approved for those specific field(s) of testing/methods listed in the table below. Clients are urged to verify the laboratory's current accreditation status for the particular field(s) of testing/Methods, since these can change due to proficiency status, suspension and/or withdrawal of accreditation.

Environmental Microbiology Laboratory Accreditation Program (EMLAP)

Initial Accreditation Date: 02/01/2013

EMLAP Scope Category	Field of Testing (FOT)	Component, parameter or characteristic tested	Method	Method Description *(for internal methods only)*
Bacterial	Air - Culturable	Bacterial Isolates	EM-BT-S-1221	Speciation of Aerobic Bacteria utilizing the MIDI Sherlock Instant FAME System
Bacterial	Air - Culturable	Bacterial Isolates	EM-BT-S-1576	Tests to Aid in Bacterial Identification
Bacterial	Air - Culturable	Bacterial Isolates	EM-BT-S-1583	Bacterial and Yeast Identification Using The API® System
Bacterial	Air - Culturable	Viable Impaction Samples	EM-BT-S-1051	Enumeration and Gram Stain Identification of Aerobic Bacteria in Air
Bacterial	Air - Culturable	Viable Impaction Samples	EM-USP-S-2081	USP 797 Bacteria and Fungi Analysis, Air Plates
Bacterial	Bulk - Culturable	Bacterial Isolates	EM-BT-S-1221	Speciation of Aerobic Bacteria utilizing the MIDI Sherlock Instant FAME System
Bacterial	Bulk - Culturable	Bacterial Isolates	EM-BT-S-1576	Tests to Aid in Bacterial Identification
Bacterial	Bulk - Culturable	Bacterial Isolates	EM-BT-S-1583	Bacterial and Yeast Identification Using The API® System
Bacterial	Bulk - Culturable	Dust, Swab, Bulk, Water/Liquids, Wipes, Contact Plates	EM-BT-S-1050	Enumeration and Gram Stain Identification of Aerobic Bacteria and Thermophilic Actinomycetes in Contact Plates, Swab, Bulk, CarpetChek and Water Samples

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EMLAP Scope Category	Field of Testing (FOT)	Component, parameter or characteristic tested	Method	Method Description *(for internal methods only)*
Bacterial	Bulk - Culturable	Dust, Swab, Bulk, Water/Liquids, Wipes, Contact plates, etc.	EM-PR-S-1040	Preparation of Bulk, Dust/ Soil, Swab/Wipe and Water/Liquid Samples for Quantitative Fungal and /or Bacterial Analysis
Bacterial	Bulk - Culturable	Gloved fingertip samples	EM-USP-S-2080	USP 797 Gloved Fingertip Sample Analysis
Bacterial	Bulk - Culturable	Media fill test vials	EM-USP-S-2083	USP 797 Media Fill Test Analysis
Bacterial	Legionella	Water, Swabs	EM-BT-S-1045	Detection and Enumeration of Legionella bacteria (based on ISO 11731:2017 Method)
Bacterial	Legionella	Water, Swabs, Wipes, Bulk, Air	EM-BT-S-1687	Detection and Enumeration of Legionella bacteria (based on CDC method)
Bacterial	Surface - Culturable	Bacterial Isolates	EM-BT-S-1221	Speciation of Aerobic Bacteria utilizing the MIDI Sherlock Instant FAME System
Bacterial	Surface - Culturable	Bacterial Isolates	EM-BT-S-1576	Tests to Aid in Bacterial Identification
Bacterial	Surface - Culturable	Bacterial Isolates	EM-BT-S-1583	Bacterial and Yeast Identification Using The API® System
Bacterial	Surface - Culturable	Dust, Swab, Bulk, Water/Liquids, Wipes, Contact Plates	EM-BT-S-1050	Enumeration and Gram Stain Identification of Aerobic Bacteria and Thermophilic Actinomycetes in Contact Plates, Swab, Bulk, CarpetChek and Water Samples
Bacterial	Surface - Culturable	Dust, Swab, Bulk, Water/Liquids, Wipes, Contact plates, etc.	EM-PR-S-1040	Preparation of Bulk, Dust/ Soil, Swab/Wipe and Water/Liquid Samples for Quantitative Fungal and /or Bacterial Analysis
Bacterial	Surface - Culturable	Surface (swab & contact plate)	EM-USP-S-2082	USP 797 Bacteria and Fungi Analysis, Surface Swabs and Contact Plates
Fungal	Air - Culturable	Viable Impaction Samples	EM-MY-S-1043	Preparation and Analysis of Air Samples for Culturable Fungi
Fungal	Air - Culturable	Viable Impaction Samples	EM-USP-S-2081	USP 797 Bacteria and Fungi Analysis, Air Plates
Fungal	Air - Direct Examination	Spore Trap Air Samples	EM-MY-S-1038	Preparation and Analysis of Spore Trap (Air) Samples for Fungal Spores, Other Biological and Non-Biological Particles
Fungal	Bulk - Culturable	Dust, Swab, Bulk, Water/Liquids, Wipes	EM-MY-S-1040	Preparation of Bulk, Dust/ Soil, Swab/Wipe and Water/Liquid Samples for Quantitative Fungal and /or Bacterial Analysis

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EMLAP Scope Category	Field of Testing (FOT)	Component, parameter or characteristic tested	Method	Method Description *(for internal methods only)*
Fungal	Bulk - Culturable	Dust, Swab, Bulk, Water/ Liquids, Wipes, etc.	EM-MY-S-2584	Analysis of Dust, Swab, Water, and Bulk Samples for Culturable Fungi
Fungal	Bulk - Direct Examination	Tape, Swab, Wipe, Bulk, Dust, Soil	EM-MY-S-1039	Preparation and Analysis of Tape, Swab, Wipe, Bulk and Dust - Soil Samples for Qualitative Direct Microscopic Examination
Fungal	Bulk - Direct Examination	Tape, Swab, Wipe, Bulk, Dust, Soil	EM-MY-S-1041	Preparation and Analysis of Tape, Swab, Wipe, Bulk and Dust - Soil Samples for Quantitative Direct Microscopic Examination
Fungal	Surface - Culturable	Dust, Swab, Bulk, Water/Liquids, Wipes	EM-MY-S-1040	Preparation of Bulk, Dust/ Soil, Swab/Wipe and Water/Liquid Samples for Quantitative Fungal and /or Bacterial Analysis
Fungal	Surface - Culturable	Dust, Swab, Bulk, Water/ Liquids, Wipes, etc.	EM-MY-S-2584	Analysis of Dust, Swab, Water, and Bulk Samples for Culturable Fungi
Fungal	Surface - Culturable	Surface (swab & contact plate)	EM-USP-S-2082	USP 797 Bacteria and Fungi Analysis, Surface Swabs and Contact Plates
Fungal	Surface - Direct Examination	Tape, Swab, Wipe, Bulk, Dust, Soil	EM-MY-S-1039	Preparation and Analysis of Tape, Swab, Wipe, Bulk and Dust - Soil Samples for Qualitative Direct Microscopic Examination
Fungal	Surface - Direct Examination	Tape, Swab, Wipe, Bulk, Dust, Soil	EM-MY-S-1041	Preparation and Analysis of Tape, Swab, Wipe, Bulk and Dust - Soil Samples for Quantitative Direct Microscopic Examination
Molecular	RT-PCR - Legionella pneumophila SG-1/ All serogroups and Legionella spp.	Water, Swabs	EM-BC-S-8678	PCR detection and quantification of Legionella 3-plex Panel in water and swab samples
Molecular	RT-PCR - Legionella spp. and Legionella pneumophila	Water, Swabs	EM-BC-S-8452	PCR detection and quantification of Legionella spp. and Legionella pneumophila in water and swab samples using BioRad iQCheck® Legionella technology
Molecular	RT-PCR - SARS-CoV-2	Air Cassettes	EM-BC-S-8625	Quantitative Detection of Coronavirus on Air Samples (RT-PCR)
Molecular	RT-PCR - SARS-CoV-2	Swabs, Air Cassettes, and Other Surface Sampling Devices	EM-BC-S-8576	Detection of Coronavirus on Surface and Air Samples (RT-PCR)

A complete listing of currently accredited EMLAP laboratories is available on the AIHA LAP, LLC website at: http://www.aihaaccreditedlabs.org